Renal Cystic Disease and Ammoniagenesis in Han:SPRD Rats^{1,2}

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ABSTRACT

Cyst formation in conditions associated with increased renal ammoniagenesis (hypokalemia, distal renal tubular acidosis, renal mass reduction) and experimental links between increased ammoniagenesis and interstitial inflammation have suggested a role for ammonia in the pathogenesis of polycystic kidney disease (PKD). To explore this hypothesis, Han: SPRD rats, a PKD model that affects male more severely than female animals, have been used. Heterozygous cystic (Cy/+) and homozygous normal (+/+) male and female offspring of Cy/+ rats were divided at 3 wk of age into control groups drinking water and experimental groups drinking 300 mM NH₄CI, 300 mM KHCO₃, 200 mM KHCO₃, 200 mM KCI, 200 mM NaHCO3, or 200 mM NaCl. At 2 months of age, the rats were kept fasting from 8:00 p.m. to 8:00 a.m. in metabolic cages and urine samples were collected under mineral oil. The rats were then weighed and anesthetized for the collection of blood and kidneys. The administration of 300 mM NH₄Cl. and to a lesser extent that of 200 mM NaCl, was accompanied by an increase in the urinary excretion of ammonia and aggravation of the renal cystic disease. On the other hand, the administration of 300 mM KHCO₃, 200 mM KHCO₃, or 200 mM NaHCO₃ lowered the urinary excretion of ammonia and markedly reduced the severity of the cystic disease and interstitial inflammation. The administration of 300 mM KHCO₃, and to a lesser extent that of 200 mM KHCO₃, resulted in the precipitation of calcium phosphate in the medullary collecting ducts. These observations are consistent with the hypothesis that renal ammoniagenesis or the metabolic processes linked to it play a role in the pathogenesis of PKD and demonstrate a

protective effect of alkali administration on the development of cystic disease in Han:SPRD rats.

Key Words: Autosomal dominant polycystic kldney disease, renal ammoniagenesis, acidosis, alkalosis, Han:SPRD rats

olycystic kidney disease has a complex pathogenesis that includes abnormalities in the proliferation of the tubular epithelial cells, fluid secretion, and remodeling of the extracellular matrix (1-6). The possibility that ammonia is involved in the pathogenesis of polycystic kidney disease was suggested (7) after the observation of an association between chronic hypokalemia and acquired renal cysts (8,9). Acquired renal cysts have also been observed in chronic renal failure, both clinically (10-12) and experimentally (13), and in patients with distal renal tubular acidosis (14,15). Increased renal ammoniagenesis, either in absolute terms or relative to the number of surviving nephrons (16-18), is common to these conditions. Abnormalities in the urinary excretion of ammonia have been described in autosomal dominant polycystic kidney disease (ADPKD) (19,20). To investigate a possible link between alterations in renal ammoniagenesis and the development of renal cystic disease, we have used Han: SPRD rats, a recently characterized model of ADPKD (21,22).

METHODS

Experimental Animals

Han:SPRD rats were obtained from the polycystic kidney program at the University of Kansas Medical Center. The animals used in this study were the offspring from heterozygous rats. The rats with homozygous disease (Cy/Cy) were recognized at 1 wk of age by the marked renal enlargement, died of uremia at 3 to 4 wk of age, and were not used in this study. The remaining homozygous normal (+/+) and heterozygous diseased (Cy/+) animals were divided into different experimental groups designed to alter ammonia production or to control for the administration of sodium or potassium. The severity of the cystic disease differs in male and female Cy/+ rats. Impairment of renal function is noticeable by 8 wk of age in male Cy/+ rats, whereas female Cy/+ rats have a milder disease (22).

Experimental Groups

At 3 wk of age, Cy/+ and +/+ Han:SPRD rats were randomly divided into control groups drinking water and experimental groups drinking 300 mM NH₄Cl, 300 mM KHCO₃, 200 mM KHCO₃, 200 mM NaHCO₃, 200 mM KCl, or 200 mM NaCl. Only female rats, which have milder renal cystic disease, received 300 mM NH₄Cl, and only male rats, which have more severe renal cystic disease, were given 300 mM KHCO₃. All rats were fed a standard rodent diet containing 23% protein (Purina Mills Inc., Richmond, IN).

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Experimental Protocol

At 2 months of age, the rats were placed in metabolic cages and kept fasting from 8 p.m. to 8 a.m. for the collection of urine under mineral oil. After the completion of the 12-h urine collection, the rats were weighed and anesthetized with Inactin (Promonta, Hamburg, Germany), 100 mg/kg body wt ip. Heparinized blood samples were obtained by cardiac puncture, the abdomen was opened, and the kidneys were removed, placed in preweighed containers with 4% paraformaldehyde, weighed, fixed overnight at 4°C, and embedded in paraffin for histologic studies.

Laboratory Methods and Morphologic Analysis

Plasma and urine creatinine concentrations were measured by an adaptation of the Jaffe reaction to an automatic chemical analyzer (23). Urinary ammonia was measured by the Berthelot method (24). Four-micrometer transverse tissue sections including cortex, medulla, and papilla were stained with hematoxylin and eosin and von Kossa stains (25). These sections were graded without knowledge of group assignment as to the extent of the cystic changes (0, absence of cysts; 1, 2, 3, and 4, cysts in <20%, 20 to 40%, 40 to 60% and >60% of renal cortex, respectively) and to the extent and severity of the interstitial infiltration by inflammatory cells (0, absence of infiltrates; 1, focal, mild; 2, focal, moderate; 3, diffuse, mild; 4, diffuse, moderate or severe).

Statistical Analysis

Comparisons of the means between control and experimental groups were made by use of the t test. All P values reported are two tailed, and the conventional cutoff of 0.05 was taken to reflect statistical significance.

RESULTS

The weights, concentrations of plasma bicarbonate, urinary excretions of ammonia, and creatinine clearances of the male and female Cy/+ control and experimental rats are summarized in Table 1. Male Cy/+

rats drinking 300 or 200 mM KHCO₃ and female Cy/+ rats drinking 300 mM NH₄Cl or 200 mM NaCl had retarded growth as compared with the control animals. Plasma bicarbonate concentrations were significantly lower in the groups drinking 300 mM NH₄Cl or 200 mM NaCl and in female rats given 200 mM KCl and were significantly higher in the male rats drinking KHCO₃ or NaHCO₃ and in the female animals given NaHCO3. Urinary excretions of ammonia were markedly increased in the rats drinking NH4Cl and to a lesser extent in those drinking NaCl and were significantly reduced in the groups treated with KHCO₃ or NaHCO₃. No significant changes in the urinary excretion of ammonia were detected in the groups receiving 200 mM KCl. Creatinine clearances were significantly reduced in the 300 mM NH₄Cl group. No significant differences in creatinine clearance were detected between the control and the remaining experimental groups.

Because of the small number of +/+ rats (22 male and 19 female animals divided in 2 control and 10 experimental groups), comparisons between individual Cy/+ and +/+ groups were limited (results not shown). The weights of Cy/+ rats receiving 300 mM KHCO₃, 200 mM KHCO₃, or 300 mM NH₄Cl were significantly lower than those of +/+ rats. The concentrations of plasma bicarbonate of male control and female NH₄Cl Cy/+ rats were significantly lower than those of +/+ rats. No significant differences in the urinary excretion of ammonia were detected between Cy/+ and +/+ rats. Creatinine clearances of male Cy/+ rats in the control and 200 mM NaCl groups and of female Cy/+ rats in the 300 mM NH₄Cl group were significantly reduced as compared with those of +/+ rats.

TABLE 1. Characteristics of male and female Cy/+ Han:SPRD rats in the control and experimental groups^a

Rat	N	Weight (g)	Plasma Bicarbonate (mEq/L)	Urine NH₄ (µmol/h per 100 g body wt)	Creatinine Clearance (mL/min per 100 g body wt)	
Male	•					
Control	5	237 ± 39	21.3 ± 0.5	9.7 ± 2.3	0.87 ± 0.31	
KHCO ₃ (300 mM)	7	156 ± 22 ^b	24.5 ± 1.7 ^b	4.5 ± 1.5 ^b	1.25 ± 0.31	
KHCO ₃ (200 mM)	5	191 ± 21 ⁶	23.7 ± 0.6^{b}	5.2 ± 0.7 ^b	1.09 ± 0.14	
NaHCO ₃ (200 mM)	5	263 ± 13	25.5 ± 3.3 ^b	5.0 ± 0.8^{b}	1.25 ± 0.20	
KCI (200 mM)	8	229 ± 25	21.4 ± 1.2	12.9 ± 2.6	1.01 ± 0.16	
NaCl (200 mM)	4	242 ± 11	19.4 ± 0.6 ^b	16.7 ± 2.7 ^b	0.90 ± 0.26	
Female `						
Control	5	189 ± 9	23.5 ± 2.0	15.4 ± 2.0	1.16 ± 0.22	
NH ₄ CI (300 mM)	5	104 ± 38 ^b	17.7 ± 4.3 ^b	83.7 ± 44.6 ^b	0.50 ± 0.22^{b}	
KHCO ₃ (200 mM)	5	168 ± 20	23.8 ± 1.6	3.4 ± 1.1 ^b	1.00 ± 0.08	
NaHCO ₃ (200 mM)	4	192 ± 20	27.9 ± 2.9 ^b	6.0 ± 0.6 ^b	1.26 ± 0.08	
KCI (200 mM)	5	182 ± 6	20.9 ± 1.0 ^b	15.2 ± 3.8	1.03 ± 0.29	
NaCi (200 mM)	6	167 ± 8 ^b	20.4 ± 1.9 ^b	27.2 ± 7.2^{b}	0.80 ± 0.33	

^a Values are mean ± SD.

 $^{^{\}rm b}$ P < 0.05 as compared with control.

The relative kidney weights and histologic scores of male and female Cy/+ are shown in Table 2. Males had significantly higher relative kidney weights than did females. The administration of 300 mM NH₄Cl to female Cy/+ rats resulted in marked renal enlargement. The administration of 200 mM NaCl to male and female Cy/+ rats caused renal enlargement of a lesser degree than that observed after the administration of 300 mM NH₄Cl. On the other hand, the administration of 300 mM KHCO₃, 200 mM KHCO₃, and 200 mM NaHCO₃ consistently resulted in a marked reduction in the size of the kidneys. The administration of 200 mM KCl had no significant effect. For the purpose of comparison, the relative kidney weights of male and female +/+ rats in the control group were 0.81 ± 0.02 and 0.78 ± 0.04 g/100 g body wt. The administration of 300 mM NH₄Cl to female +/+ rats and of 300 mM KHCO₃ to male +/+ rats was accompanied by a slight increase in relative kidney weight (results not shown).

Macroscopic and microscopic examinations of the kidneys confirmed the marked aggravation of renal cystic disease caused by the administration of 300 mM NH₄Cl (Figures 1 to 3; Table 2) and to a lesser extent by the administration of 200 mM NaCl, as well as the striking protective effect afforded by the administration of 200 or 300 mM KHCO3 or 200 mM NaHCO₃ (Figures 4 to 6; Table 2). The administration of KHCO₃ or NaHCO₃ was accompanied by a marked reduction not only in the number of cysts, but also in the density of cellular infiltrates in the interstitium (Figure 6). The administration of 300 mM KHCO₃ was accompanied by the intraluminal deposition of calcium phosphate in the medulla, which was more marked in Cy/+ rats than in +/+ rats (Figure 7). The intraluminal precipitation of calcium phosphate was less prominent in the rats drinking 200 mM KHCO₃ and was not observed in the animals receiving 200 mM NaHCO₃.

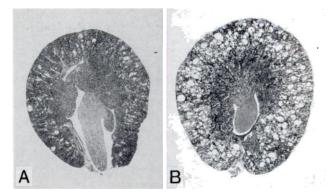


Figure 1. Representative transverse sections of kidneys from female Cy/+ Han:SPRD rats from the control group (A) or from the group drinking 300 mM NH₄Cl (B). Note the marked aggravation of the renal cystic disease observed in the rat drinking 300 mM NH₄Cl.

DISCUSSION

This study clearly shows that the development of inherited renal cystic disease can be markedly altered by environmental or dietary factors. Although the rats were not pair fed, the observations of this study cannot be explained by different caloric or protein intakes. Interventions causing similar degrees of growth retardation had opposite effects on the development of renal cystic disease, whereas consistent results were obtained by interventions that did not cause any growth retardation. Dietary changes accompanied by reduced urinary excretions of ammonia markedly attenuated the development of renal cystic disease, whereas those accompanied by increased urinary excretions of ammonia had the opposite effect. Thus, this study also supports the hypothesis that the ammonia or metabolic processes linked to renal ammoniagenesis may play a role in the pathogenesis of

TABLE 2. Relative kidney weights and histologic scores of male and female Cy/+ Han:SPRD rats in the control and experimental groups^a

Rat	N	Kidney/Body Wt × 100	Cystic Dilation	Interstitial Inflammation
Male				
Control	5	2.10 ± 0.27	2.80 ± 0.45	2.80 ± 0.45
KHCO ₃ (300 mM)	7	1.48 ± 0.23 ^b	1.43 ± 0.53 ^b	0.86 ± 0.38^{b}
KHCO ₃ (200 mM)	5	1.19 ± 0.12 ^b	1.20 ± 0.45 ^b	1.20 ± 0.45 ^b
NaHCO ₃ (200 mM)	5	1.20 ± 0.10 ^b	1.40 ± 0.55 ^b	1.20 ± 0.45^{b}
KCI (200 mM)	8	1.95 ± 0.16	2.63 ± 0.52	2.38 ± 0.74
NaCl (200 mM)	4	2.95 ± 0.13^{b}	3.25 ± 0.29	2.88 ± 0.25
Female				
Control	5	1.39 ± 0.11	1.80 ± 0.45	1.20 ± 0.45
NH ₄ CI (300 mM)	5	2.81 ± 0.51 ^b	3.40 ± 0.55^{b}	2.10 ± 0.22^{b}
KHCO ₃ (200 mM)	5	1.13 ± 0.07 ^b	1.10 ± 0.22 ^b	0.80 ± 0.27
NaHCO ₃ (200 mM)	4	1.13 ± 0.07 ^b	1.10 ± 0.22 ^b	0.80 ± 0.45
KCI (200 mM)	5	1.50 ± 0.07	1.80 ± 0.45	1.20 ± 0.45
NaCl (200 mM)	6	2.20 ± 0.31^{b}	2.50 ± 0.45^{b}	1.83 ± 0.41^{b}

^a Values are mean ± SD.

 $^{^{\}rm b}$ P < 0.05 as compared with control.

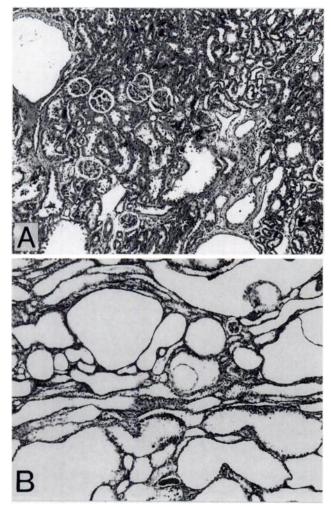


Figure 2. Four-micrometer kidney sections from female Cy/+ Han:SPRD rats from the control group (A) or from the group drinking 300 mM NH₄Cl (B). Note the extensive cystic disease in the rat drinking 300 mM NH₄Cl. Hematoxylin and eosin, \times 100.

polycystic kidney disease. Consistent with these results are the observations by Cowley *et al.* of worse renal cystic disease in Han:SPRD Cy/+ rats fed NH₄Cl or a potassium-deficient diet (26).

The use of alkali in the treatment of renal diseases is not new. It was recommended by Richard Bright, and it was an accepted therapy for many years (27). A number of experimental animal studies at the turn of the century claimed that the administration of alkali could partially prevent the nephrotoxic effects of certain anesthetics (28) or metallic compounds such as uranium nitrate (29,30). In a prospective study of patients with scarlet fever, it was found that the prophylactic administration of large amounts of sodium bicarbonate and potassium citrate reduced the frequency of scarlatinal nephritis (31). The protective effect of alkali administration has been confirmed many years later in a number of renal conditions

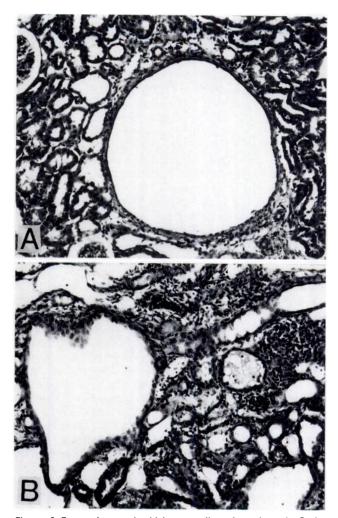


Figure 3. Four-micrometer kidney sections from female Cy/+ Han:SPRD rats from the control group (A) or from the group drinking 300 mM NH₄Cl (B). Note the presence of inflammatory cell infiltrates in the rat drinking 300 mM NH₄Cl. Hematoxylin and eosin, $\times 200$.

including subtotal nephrectomy (32) and hypokalemic nephropathy (33). The results of our study indicate that the autosomal dominant model of renal cystic disease in Han:SPRD rats is also markedly susceptible to changes in acid base balance and that pathologic changes can be markedly attenuated by the administration of alkali.

Insufficient understanding of the mechanisms by which changes in acid base metabolism could affect the development of renal disease has likely inhibited the interest of clinicians in this potential form of therapy. Common to many renal diseases, including those where the administration of alkali has been shown to be protective, as well as many renal cystic diseases, is the presence of interstitial cellular infiltrates and fibrosis. Because free base ammonia can activate complement by disrupting a reactive internal thioester bond within the alpha subunit of the third

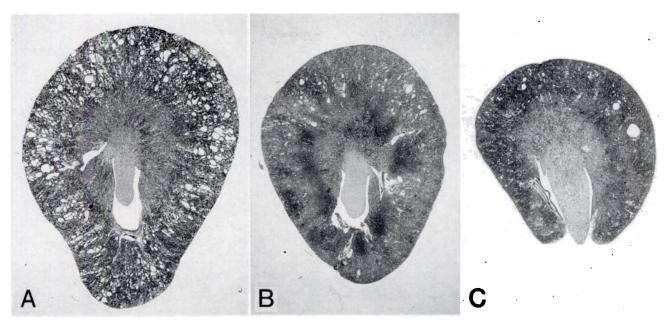


Figure 4. Representative transverse sections of kidneys from male Cy/+ Han:SPRD rats from the control group (A), from the group drinking 200 mM NaHCO₃ (B), or from the group drinking 200 mM KHCO₃ (C). Note that the severity of the cystic disease is much less in the rats drinking 200 mM NaHCO₃ or 200 mM KHCO₃.

component of complement (34), it has been proposed that the enhanced cortical production of ammonia associated with renal mass reduction (32), chronic hypokalemia (33), and the dietary deficiency of antioxidants (35) may be responsible for the development of interstitial inflammation and fibrosis in these conditions. A similar mechanism may be operative in polycystic kidney disease because patients with this disease may have a defect in the transfer of ammonia to the final urine (20) analogous to that observed after subtotal nephrectomy (36).

The results of our study indicate that the administration of acids and alkalis has a marked effect not only on the development of interstitial inflammation, but also on cyst formation. Alkali administration was previously found to reduce cystic tubular dilation in chronic hypokalemic nephropathy and after subtotal nephrectomy (32.33). The enhanced renal production of ammonia could be linked to cyst formation by a number of mechanisms. The local generation of autacoids, cytokines, and growth factors as a result of the ammonia-induced complement activation and inflammation in the renal interstitium may contribute to abnormal growth and/or fluid secretion by the tubular epithelium. In nonrenal cells, ammonia can stimulate DNA (37), RNA, and protein synthesis (37,38) and decrease the rate of protein (39-41) and glycosaminoglycan (42,43) degradation. In rabbit proximal tubular cells, ammonia results in an increase in RNA and protein content, stimulation of protein synthesis, and inhibition of protein degradation, without change in DNA synthesis (44,45). Finally, metabolic factors linked to renal ammoniagenesis, rather than to ammonia per se, may be important. Glutamine oxidation by the mitochondrial phosphate-dependent glutaminase pathway is an important source of ATP in the proximal tubular epithelial cells (46,47). Extracellular ATP is a mitogen for a number of mammalian cells (48-52), and mitochondrial drugs that deplete the ATP content can result in growth inhibition and cell differentiation (53,54). The addition of exogenous adenine nucleotides to isolated rabbit kidney tubules enriched in proximal segments increases the cell content of ATP (55), and the mitogenic effect of adenine nucleotides on proximal tubular epithelial cells greatly exceeds that of other growth-promoting agents (56,57). We have also found that ATP has a strong mitogenic effect on cyst-derived epithelial cell cultures (V.E. Torres, D.K. Mujwid, unpublished observation).

It is uncertain to what extent the observations in this study are relevant to human renal cystic disease. Nevertheless, certain observations are consistent with a role for renal ammoniagenesis or a metabolic process linked to renal ammoniagenesis in the pathogenesis of acquired renal cystic disease and ADPKD. Studies in animal models with reduced renal mass and in humans with chronic renal disease have shown that, although the absolute excretion of ammonia is reduced, the excretion of ammonia per nephron unit is increased (16-18). In the remnant kidney model, a defective trapping in the renal medulla causes a reduction in the urinary excretion of ammonia, despite an enhanced production and concentration in the renal cortex (36). It seems likely that the high production and concentration of ammonia also occur in the surviving nephrons of dialysis patients because stand-

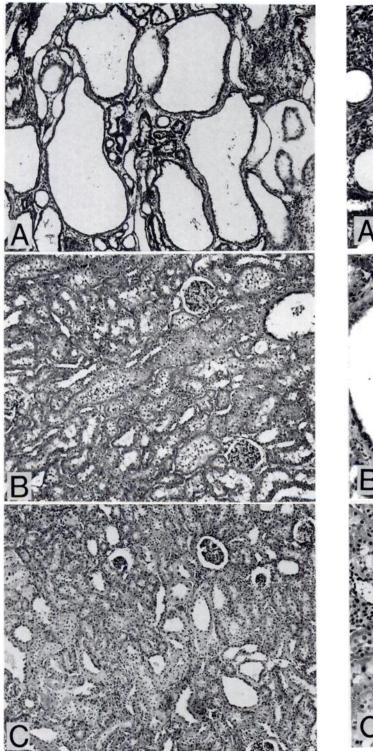


Figure 5. Four-micrometer kidney sections from male Cy/+ Han:SPRD rats from the control group (A), from the group drinking 200 mM NaHCO $_3$ (B), or from the group drinking 200 mM KHCO $_3$ (C). Note that the severity of the cystic disease is much less in the rats drinking 200 mM NaHCO $_3$ or 200 mM KHCO $_3$. Hematoxylin and eosin, $\times 100$.

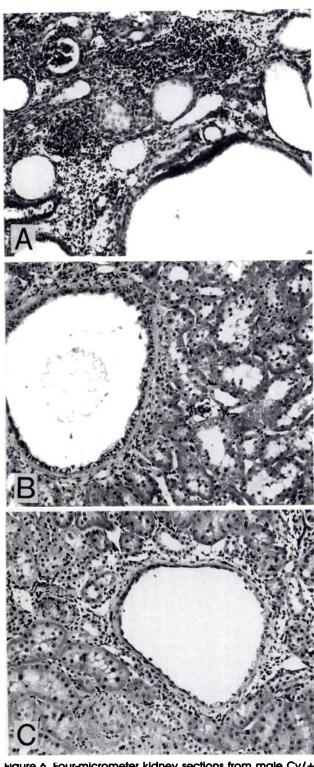


Figure 6. Four-micrometer kidney sections from male Cy/+ Han:SPRD rats from the control group (A), from the group drinking 200 mM NaHCO $_3$ (B), or from the group drinking 200 mM KHCO $_3$ (C). Note that the severity of the inflammatory cell infiltrates is much less in the rats drinking 200 mM NaHCO $_3$ or 200 mM KHCO $_3$.

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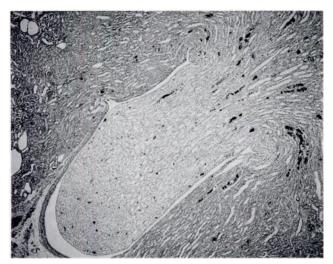


Figure 7. Four-micrometer tissue section from a male Cy/+ Han:SPRD rat drinking 300 mM KHCO $_3$. Note the mild renal cystic disease and the presence of extensive precipitation of calcium phosphate in the medullary collecting ducts. von Kossa, $\times 25$.

ard bicarbonate and acetate dialysates do not completely correct uremic acidosis (58). In ADPKD, the disruption of the normal corticomedullary vascular-tubular architecture by cysts is likely responsible for the renal concentration defect that is the earliest functional abnormality in this disease (59). ADPKD patients with normal GFR cannot transfer ammonia normally to the urine, likely also because of the loss of the corticomedullary concentration gradient (20). To compensate for this transport defect, the cortical production of ammonia may be increased in ADPKD patients at an earlier stage of renal insufficiency and contribute to the progression of the disease.

Note added in proof: Additional experiments have been performed to determine the effects of lower concentrations of NaHCO₃. The administration of 75, 150, and 200 mM NaHCO₃ significantly reduced the increase in renal size of cystic over that of nonaffected rats by 43, 62, and 70%, respectively.

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REFERENCES

- Grantham JJ: Fluid secretion, cellular proliferation, and the pathogenesis of renal epithelial cysts. J Am Soc Nephrol 1993;3:1843–1857.
- Ye M, Grantham JJ: The secretion of fluid by renal cysts from patients with autosomal dominant polycystic kidney disease. N Engl J Med 1993;329:310-313.
- Wilson PD, Hreniuk D, Gabow PA: Abnormal extracellular matrix and excessive growth of human adult polycystic kidney disease epithelia. J Cell Physiol 1992;150: 360-369.
- Wilson PD, Sherwood AC, Palla K, Du J, Watson R, Norman JT: Reversed polarity of Na+-ATPase: Misloca-

- tion to apical plasma membranes in polycystic kidney disease epithelia. Am J Physiol 1991;260:F420-F430.
- Carone FA, Nakamura S, Punyarit P, Kanwar YS, Nelson WJ: Sequential tubular cell and basement membrane changes in polycystic kidney disease. J Am Soc Nephrol 1992;3:244-253.
- Avner ED, Sweeney WE Jr, Ellis D: In vitro modulation of tubular cyst regression in murine polycystic kidney disease. Kidney Int 1989;36:960-968.
- Alpern RJ, Toto RD: Hypokalemic nephropathy—a clue to cystogenesis? N Engl J Med 1990;322:398–399.
- Torres VE, Young WF Jr, Offord KP, Hattery RR: Association of hypokalemia, aldosteronism, and renal cysts. N Engl J Med 1990;322:345–351.
- Liddle GW, Bledsoe T, Coppage WS Jr: A familial renal disorder simulating primary aldosteronism but with negligible aldosterone secretion. Trans Assoc Am Physicians 1963;76:199-213.
- Matson MA, Cohen EP: Acquired cystic kidney disease: occurrence, prevalence, and renal cancers. Medicine 1990;69:217-226.
- 11. Ishikawa I: Uremic acquired renal cystic disease: Natural history and complications. Nephron 1991;58:257-
- Levine E, Slusher SL, Grantham JJ, Wetzel LH: Natural history of acquired renal cystic disease in dialysis patients: A prospective longitudinal CT study. AJR Am J Roentgenol 1991;156:501–506.
- Kenner CH, Evan AP, Blomgren P, Aronoff GR, Luft FC: Effect of protein intake on renal function and structure in partially nephrectomized rats. Kidney Int 1985;27: 739-750.
- Morin D, Picou G, Dumas R: Clinical quiz. Chronic potassium deficiency. Pediatr Nephrol 1991;5:669-670.
- Igarashi T, Shibuya K, Kamoshita S, et al.: Renal cyst formation as a complication of primary distal renal tubular acidosis. Nephron 1991;59:75-79.
- Tannen RL: Renal ammonia production and excretion. In: Windhager EE, Ed. Handbook of Physiology. Section 8. Renal Physiology. Vol. I. 1992:1017–1059.
- 8. Renal Physiology. Vol. I. 1992:1017–1059.

 17. Halperin ML, Kamel KS, Ethier JH, Stinebaugh BJ, Jungas RL: Blochemistry and physiology of ammonium excretion. In: Seldin DW, Giebisch G, Eds. The Kidney: Physiology and Pathophysiology. 2nd Ed. New York: Raven Press, Ltd; 1992:2645–2680.
- Donnelly S, Kamel KS, Vasuvattakul S, Narins RG, Halperin ML: Might distal renal tubular acidosis be a proximal tubular cell disorder? Am J Kidney Dis 1992; 19:272-281.
- Preuss H, Geoly K, Johnson M, Chester A, Kliger A, Schreiner G: Tubular function in adult polycystic kidney disease. Nephron 1979;24:198–204.
- Torres VE, Keith DS, Offord KP, Kon SP, Wilson DM: Renal ammonia in autosomal dominant polycystic kidney disease. Kidney Int 1994;45:1745–1753.
- Kaspareit-Rittinghausen J, Rapp K, Deerberg F, Wcislo A, Messow C: Hereditary polycystic kidney disease associated with osteorenal syndrome in rats. Vet Pathol 1989;26:195-201.
- Cowley BD Jr, Gudapaty S, Kraybill AL, et al.: Autosomal dominant polycystic kidney disease in the rat. Kidney Int 1994;43:522–534.
- Chasson AL, Grady HJ, Stanley MA: Determination of creatinine by means of automatic chemical analysis. Am J Clin Pathol 1961;35:83–88.
- Tietz NW, Ed. Fundamentals of Clinical Chemistry. 3rd
 Ed. Philadelphia: WB Saunders Company; 1987:748–749.
- Sheehan DC, Hrapchak BB: Theory and Practice of Histopathology. 2nd Ed. St. Louis: CV Mosby Co; 1980.
- Cowley BD Jr, Grantham JJ, Muessel MJ, Gattone VH II: Accelerated progression of inherited polycystic kidney disease (PKD) caused by non-genetic interventions [Abstract 73P]. J Am Soc Nephrol 1993;4:261.
- 27. Osman AA: Studies in Bright's disease. The use of alkalies in the treatment of Bright's disease, and their prophylactic value in this and other conditions associ-

- ated with impairment of renal function. Guy's Hosp Rep 1927;77:386-435.
- MacNider WB: The stability of the acid-base equilibrium
 of the blood in naturally nephropathic animals and the
 effect on renal function of changes in this equilibrium. J
 Exp Med 1918:28:517-528.
- Exp Med 1918;28:517-528.

 29. MacNider WB: The inhibition of the toxicity of uranium nitrate by sodium carbonate, and the protection of the kidney acutely nephropathic from uranium from the toxic action of an anesthetic by sodium carbonate. J Exp Med 1916;23:171-187.
- Goto K: A study of the acidosis, blood urea, and plasma chlorides in uranium nephritis in the dog, and of the protective action of sodium carbonate. J Exp Med 1917; 25:693-719.
- Carter H, Osman AA: The prevention of scarlatinal nephritis. Proc R Soc Med 1927;20:1405–1408.
- Nath KA, Hostetter MK, Hostetter TH: Pathophysiology of chronic tubulo-interstitial disease in rats. Interactions of dietary acid load, ammonia, and complement component C3. J Clin Invest 1985;76:667–675.
- 33. Tolins J, Hostetter M, Hostetter T: Hypokalemic nephropathy in the rat: The role of ammonia in chronic tubular injury. J Clin Invest 1987:79:1447-1458.
- tubular injury. J Clin Invest 1987;79:1447-1458.

 34. Hostetter MK, Johnson GM, Retsinas EM: The erythrocyte as instigator of inflammation. Generation of amidated C3 by erythrocyte adenosine deaminase. J Clin Invest 1989;84:665-671.
- 35. Nath CA, Salahudeen AK: Induction of renal growth and injury in the intact rat kidney by dietary deficiency of antioxidants. J Clin Invest 1990;86:1179-1192.
 36. Buerkert J, Martin D, Trigg D, Simon E: Effect of reduced renal mass on ammonium handling and net
- Buerkert J, Martin D, Trigg D, Simon E: Effect of reduced renal mass on ammonium handling and net acid formation by the superficial and juxtamedullary nephron of the rat. J Clin Invest 1983;71:1661-1675.
 Dubé F, Epel D: The relationship between intracellular
- 37. Dubé F, Epel D: The relationship between intracellular pH and rate of protein synthesis in sea urchin eggs and the existence of a pH-independent event triggered by ammonia. Exp Cell Res 1986;162:191-204.
 38. Shiokawa K, Kawazoe Y, Nomura H, et al.: Ammonium
- Shiokawa K, Kawazoe Y, Nomura H, et al.: Ammonium ion as a possible regulator of the commencement of rRNA synthesis in Xenopus laevis embryogenesis. Dev Biol 1986;115:380-391.
- Hopgood MF, Clark M, Ballard FJ: Inhibition of protein degradation in isolated rat hepatocytes. Biochem J 1977;164:399-407.
- 1977;164:399-407.
 40. Poole BS, Ohkuma S, Warbarton MJ: The accumulation of weakly basic substances in lysosomes and the inhibition of intracellular protein degradation. Acta Biol Med Geriatr 1977;36:1777-1788.
- Dean R: Protein degradation in cell cultures: General considerations on mechanisms and regulation. Fed Proc 1980;39:15-19.
- Glimelius B, Westermark B, Wasteson A: Ammonium ion interferes with the lysosomal degradation of glycosaminoglycans in cultures of human glial cells. Exp Cell Res 1977;108:23–30.
- Wiesmann UN, Colombo JP, Bachmann C: Intralysosomal generation of ammonia from urea by endocytosed

- urease results in secretion of free lysosomal ary lsulfatase-A and increased activity of membrane-bound β -glucosidase in cultured brain cells. Enzyme 1991;45:222–232.
- Golchini K, Norman J, Bohman R, Kurtz I: Induction of hypertrophy in cultured proximal tubule cells by extracellular NH₄Cl. J Clin Invest 1989;84:1767–1779.
- 45. Kurtz I: Role of ammonia in the induction of renal hypertrophy. Am J Kidney Dis 1991;17:650-653.
- Wirthensohn G, Guder WG: Renal substrate metabolism. Physiol Rev 1986;66:469-497.
- Brosnan JT, Lowry M, Vinay P, Gougoux A, Halperin ML: Renal ammonium production. Can J Physiol Pharmacol 1987;65:489-498.
- Huang N, Wang D, Heppel LA: Extracellular ATP is a mitogen for 3T3, 3T6, and A431 cells and acts synergistically with other growth factors. Proc Natl Acad Sci USA 1989;86:7904-7908.
- Schulze-Lohoff E, Zanner S, Ogilvie A, Sterzel RB: Extracellular ATP stimulates proliferation of cultured mesangial cells via P₂-purinergic receptors. Am J Physiol 1992;263:F374-F383.
- Wang D-J, Huang N-N, Heppel LA: Extracellular ATP and ADP stimulate proliferation of porcine aortic smooth muscle cells. J Cell Physiol 1992;153:221-233.
- Huang N-N, Wang D-J, Heppel LA: Stimulation of aged human lung fibroblasts by extracellular ATP via suppression of arachidonate metabolism. J Biol Chem 1993; 268:10789-10795.
- Popper LD, Batra S: Calcium mobilization and cell proliferation activated by extracellular ATP in human ovarian tumour cells. Cell Calcium 1993;14:209–218.
- 53. Fux A, Sidi Y, Kessler-Icekson G, Wasserman L, Novo-grodsky A, Nordenberg J: Dimethylthiourea inhibition of B16 melanoma growth and induction of phenotypic alterations; relationship to ATP levels. Br J Cancer 1991; 63:489–494.
- 54. Van Den Bogert C, Spelbrink JN, Dekker HL: Relationship between culture conditions and the dependency on mitochondrial function of mammalian cell proliferation. J Cell Physiol 1992;152:632–638.
- Weinberg JM, Humes HD: Increases of cell ATP produced by exogenous adenine nucleotides in isolated rabbit kidney tubules. Am J Physiol 1986;250:F720

 F733
- Kartha S, Toback FG: Purine nucleotides stimulate DNA synthesis in kidney epithelial cells in culture. Am J Physiol 1985;249:F967-F972.
- 57. Humes HD, Cieslinski DA: Adenosine triphosphate stimulates thymidine incorporation but does not promote cell growth in primary cultures of renal proximal tubule cells. Renal Physiol Biochem 1991;14:253-258.
 58. Oettinger CW, Oliver JC: Normalization of uremic acido-
- 58. Oettinger CW, Oliver JC: Normalization of uremic acidosis in hemodialysis patients with a high bicarbonate dialysate. J Am Soc Nephrol 1993;3:1804–1807.
- Gabow PA, Kaehny WD, Johnson AM, et al.: The clinical utility of renal concentrating capacity in polycystic kidney disease. Kidney Int 1989;35:675–680.

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